

RESEARCH ARTICLE

Nutritional Composition and Antibacterial Activity of Indigenous Edible Mushroom *Coprinopsis atramentaria*

CN Osuji¹, EU Nwabueze¹, TO Akunna¹ and EO Ahaotu²

¹Department of Science Laboratory Technology, Imo State Polytechnic, Umuagwo, Nigeria ²Department of Animal Production Technology, Imo State Polytechnic Umuagwo, Nigeria

ARTICLE INFO

ABSTRACT

Received: May 18, 2013 Revised: June 20, 2013 Accepted: July 12, 2013

Key words:

Antibacterial activity Coprinopsis atramentaria Proximate nutritional composition

*Corresponding Address: EO Ahaotu emmaocy@yahoo.com Edible mushrooms are the fleshy and bodies of several species of fungi. Mushrooms belong to the macro fungi because their fruiting structures are large enough to be seen with the naked eye. They can appear either below ground (hypogeous) or above ground (epigeous) where they may be picked by hand. They have been cultivated worldwide for commercial purposes. The research proves that *coprinopsis atramentaria* a nutritional mushroomand has great value to health. Antibacterial activity of various solvent extracts of *coprinopsis atramentaria* was tested against six species of bacteria: *Escherichia coli, Pseudomonas aeruginosa, Salmonella typhi, Staphylococcus aureus, Bacillussubtilis* and *Micrococcus luteus*. The methanol extract exhibited maximum activity. It showed narrow spectrum active against gram-negative bacteria and strongly inhibited gram-positivebacteria *Micrococcus luteus, Staphylococcus aureus* and *Bacillus subtilis*.

Cite This Article as: Osuji CN, EU Nwabueze, TO Akunna and EO Ahaotu, 2013. Nutritional composition and antibacterial activity of indigenous edible mushroom *Coprinopsis atramentaria*. Inter J Appl Sci Engr, 1(2): 61-65. www.ijapscengr.com

INTRODUCTION

Man has been hunting for the wild mushrooms since antiquity (Oei, 2010). During the early days of civilization, mushrooms were consumed mainly for their palatability and unique flavours (Boa, 2004). Edible mushrooms are consumed by humans as comestibles for their nutritional value and they are occasionally consumed for their supposed medicinal value. Mushrooms consumed by those practicing folk medicine are known as medicinal mushrooms. Hallucinogenic mushrooms (Psilocybin mushrooms) are occasionally consumed for recreational or religious purposes; they can produce severe nausea and disorientation and are therefore not commonly considered edible mushrooms (Arora, 1986). Present use of mushrooms is totally different from traditional because. lot of research has been done on the chemical composition of mushrooms, which revealed that mushrooms can be used as a diet to combat diseases. Many genera of mushrooms are edible and are rich inessential nutrients such as carbohydrates with low fat and oil content, proteins, vitamins, mineral, fibres and various amino acids. One third of the iron in the mushrooms is in available form (Okwulehie and Odunze, 2004). The proximate nutrient composition of indigenous edible

mushroom (Coprinopsis atramentaria) has been determined by using various laboratory analyses. The proximate nutrient contents of crude protein, crude fibre, crude fat, carbohydrate, soluble sugars, and mineral elements (Potassium, Sodium, Phosphorus, Calcium, Zinc, Iron, Copper, Manganese and Magnesium) were analysed. The analysis indicated that the indigenous edible mushroom contained rich sources of crude protein (32.23±0.89%), crude (19.7±0.09%) fibre and carbohydrate (48.55±0.21%). The protein content of mushrooms has been reported to be twice that of vegetables and four times that of oranges and significantly higher than that of wheat (Aletor, 1990; Fasidi, 1996; Okwulehie and Odunze, 2004). Mushrooms are also relatively much cheaper than beef, pork and chicken that contain similar nutrients. Due to their high content of vitamin, protein and minerals; they are considered as poor man's protein. Due to the low fat and oil content, they are recommended as good source of food supplement for patients with cardiac problems or at risk with lipidinduced disorders. They are also recommended to diabetic and anaemic persons, owing to their low carbohydrate and high folic acid content. Mattila et al. (2000) mentioned that the mushrooms like Coprinopsis atramentaria, Morchella esculenta, Helvella crispa, Hydnum

coralloides, Hypoxylon vernicosum and Polyporus mylittae were used much earlier in Asia. In general, the fruiting bodies of mushrooms contain about 56.8% carbohydrate, 25.0% protein, 5.7% fat and 12.5% ash on a dry weight basis (Ouzouni et al., 2009). Protein contents vary between 4 and 9% in Auricularia sp. and between 24 and 44% in Agaricus sp. Their energy value also varies according to species, which is about equal to that of an apple. Mushrooms also possess bioactive natural products that are antitumor, antiviral and antibacterial. They contain many different bioactive compounds with diverse biological activity depending on the way they prepared and consumed (Jordan, 2006). In recent years, multiple drug resistance in human pathogenic microorganisms has developed due to indiscriminate use of commercial antimicrobial drugs commonly used in the treatment of infectious diseases. This situation forced scientists for searching new antimicrobial substances from various sources that are the good sources of novel antimicrobial chemotherapeutic agents (Karaman et al., 2003; Rai, 1994, 1997). Bender et al. (2001) culture extracts of Coprinus comatus and other Coprinopsis atramentaria showed antibacterial activity against Staphylococcus saprophyticus. To the best of our knowledge, no research has been available on nutritional and anti-microbial activity of an indigenous edible mushroom Coprinopsis atramentaria. Therefore, the aim of the present work is to evaluate nutritional and antimicrobial potential of the test fungi.

Mushroom

The wild edible mushroom *Coprinopsis atramentaria*, indigenously collected andidentified, was cultivated under laboratory condition (Science Laboratory Technology Department, Imo State Polytechnic Umuagwo, Nigeria) to produce fruit body using compost. The mushroom cultivated and harvested was shade dried at room temperature. Dried samples were crushed by using a mortar with pestle and stored in pre-cleaned polyethylene bottles until the analysis started. Dried mushroom sample (50 g) was extracted by stirring with 500 ml of boiling water for 3 hours and filtered through Whitman No. 4 filter paper. The filtered extract was then subjected in rotor evaporator to dryness and stored at 4°C for further use.

Proximate nutritional composition

The proximate analysis (carbohydrates, fats, proteins and ash) of mushroom powder were determined by using AOAC (2000) methods. Crude protein (N \times 6.25) was determined by micro Kjeldahl method described by Pearson (1976), in which the sample was digested with a known quantity of acid. The digested material was distilled after the addition of alkali. The released ammonia was collected in 4% boric acid. The resultant boric acid, which now contained the ammonia, was then titrated against 0.1 N HCl, manually. The nitrogen content thus determined was multiplied by a factor of 6.25 to arrive at the amount of crude protein. Crude fat content of the samples was determined by extracting a known weight of powdered plant sample with petroleum ether, using by Soxhlet. The ash content was determined by combusting the mushroom material in silica crucibles in a muffle furnace at 535±150°C for 3 h. Fibre content was estimated

by the method of Van (1967). The carbohydrate content was calculated by subtracting the contents of crude ash, fat, fibre and protein from dry matter. Soluble sugars were determined as follows: The powdered samples were extracted with boiling water for30 min, absolute alcohol were added to precipitate the soluble sugars. The precipitated polysaccharides were collected bv centrifugation at 5000 rpm for 10 min in a bench centrifuge and subsequently dried at 60°C to remove residual ethanol. Soluble sugars were determined by the phenolsulphuric reaction described by Whistler and Wolfrom (1962). Minerals were determined by atomic absorption spectrophotometer (AAS-6800, Shimadzu, Japan) after drying the samples while Phosphorus was determined by using the molybdovanadate method (AOAC, 2000).

Statistical analysis

Each experiment was carried out thrice for each parameter and we had three replications (n = 3) from which we derived the mean values and standard error (SE) (Muhammed *et al.*, 2010).

Antimicrobial activity

The following strains of bacteria were used: Escherichia coli (ATCC 25992), Pseudomonas aeruginosa (ATCC27853), Salmonella typhi (clinical isolate), Staphylococcus aureus (ATCC 12598). Micrococcus luteus (clinical isolate) and Bacillus subtilis (ATCC 6633). Using the polarity difference of various solvents, dry mushroom powder (60-100 mesh) was extracted at 60°C, in order, by hexane, chloroform, methanol, ethyl acetate and boiled water. The corresponding extraction fraction for each solvent was obtained. Each extract was dissolved in dimethyl sulphoxide (DMSO) at a concentration of 50 mg/ml and stored at 4°C. Antimicrobial activity of the crude extracts was tested according to Ndyetabura (2010) by agar well assay methods as previously described by Rojas et al. (2006) and Moshi et al. (2006). Bacterial cultures were grown at 37°C for 24 h in Nutrient Broth. The culture suspensions were adjusted by comparing against 3.0 McFarland. The prepared agar plates were inoculated with 100 µl bacteria culture by spreading evenly over the surface of agar plate, using an ethanol flamed glass Drigal sky spatula (spreader). Un-inoculated on the agar, using a sterile cork-borer. For each test microorganism, 25 µl of each extract and of control were pipette into different wells (Rojas et al., 2006). The wells were then labelled to correspond with the code numbers of the test crude extracts and controls. The treated plates were stored in are frigerator at 4°C for at least 6 h to allow diffusion of the extracts into the agar while arresting the growth of the test microbes. The plates were then incubated for 24 h at 37°C. The test was carried out in triplicates. Antimicrobial activity was determined by measuring the diameters of zones of inhibition in mm using callipers. The means of the diameters of zones of growth inhibitions for the treatments were calculated and recorded.

Proximate nutritional composition

The results of the proximate and mineral composition are shown in Table 1. It shows that the dominant compounds are protein, fibre and carbohydrates and the contents are 32.23 ± 0.89 , 19.7 ± 0.09 and 48.55%, respectively. Crude fat and ash contents are 2.90 and 11.42% respectively. Of the mineral analyzed, the potassium, sodium, phosphorus, calcium content are 422 ± 0.49 , 39 ± 0.23 , 327 ± 0.29 , 81 mg/100 g and the iron, copper, manganese, zinc, magnesium content are 39 ± 0.32 , 3.72 ± 0.02 , 0.8 ± 0.01 , 1.9 ± 0.09 , $39\pm0.31 \text{ mg}/100$ g, respectively.

 Table 1: Proximate nutritional composition of Coprinopsis atramentaria

Ash (%)	11.42 ± 0.12
Fat (%)	2.90 ± 0.03
Protein % ($N \times 6.25$)	32.23 ± 0.89
Carbohydrate (%)	48.55 ± 0.21
Crude fibre (%)	19.7 ± 0.09
Sodium (mg/100 g)	39 ± 0.34
Potassium (mg/100 g)	422 ± 0.93
Calcium (mg/100 g)	81 ± 0.87
Iron (mg/100 g)	39.0 ± 0.32
Copper (mg/100 g)	3.72 ± 0.17
Zinc (mg/100 g)	1.9 ± 0.09
Magnesium (mg/100 g)	39 ± 0.31
Manganese (mg/100 g)	0.8 ± 0.05

Antimicrobial activity

The antimicrobial effect of different solvent extracts of *Coprinopsis atramentaria* was tested against three species of gram-positive bacteria, three species of gram-negative bacteria. Among the different solvent extract, methanolic extract showed more effective inhibitory activity against bacteria. As summarized in Table-2, *Coprinopsis atramentaria* methanolic extract had a narrow antibacterial spectrum against gram negative bacteria and strongly inhibited the growth of the grampositive bacteria tested, including B. *subtilis*. The maximal zones of inhibition ranged from 6 to 18 mm. The most susceptible bacterium was *M. luteus for* both boiled and raw extract with a diameter of 14 ± 1.2 and 18 ± 1.03 mm, respectively.

Against *P. aeruginosa*, both the extracts of *Coprinopsis atramentaria* showed antibacterial activity but the activity range was less in boiled extract $(2 \pm 0.25 \text{ mm})$ when compared with raw mushroom extract $(7 \pm 0.13 \text{ mm})$. This might be due to the influence of temperature that disturbed the compound, which is responsible for the activity.

DISCUSSION

Proximate compositions

In the samples, the dominant compounds were protein, fibre and carbohydrates. Some edible fungi were highly valued as a good source of protein and their protein contents usually range from 19 to 35% of dry weight, from 15.4 to26.7%, from 16.5 to 59.4% (Crisan and Sands, 1978; Yang *et al.*, 2001; Kalac, 2009). Here, protein content in *Coprinopsis atramentaria* (32.23%) seemed to have a normal protein content compared with other edible mushrooms. These relatively high protein values obtained can enrich human diet especially in villages where meat is rare and expensive. It would appear then those grams for gram of mushroom contain more protein than either potato or cabbages (can supplement

diet). The protein content of mushroom is known to be highly variable due to strain of some species, tissue type and stage development, substrate and method analysis. However, in spite of all these high values, mushrooms are still inferior in protein to such standard protein sources as meat, fish and eggs. It is also unlikely that in spite of the relatively high protein values that mushroom can serve as a sole source of protein. Crude fibre content in this mushroom (19.1%) was quite high in comparison with many other edible mushrooms (3-32%) (Crisan and Sands, 1978; Mau et al., 2002). The fairly high level of fibre in the mushroom was a desirable characteristic since fibre plays an important role in human diet. Obviously, the fruit bodies of Coprinopsis atramentaria were a good source of fibre. Carbohydrate content of Coprinopsis atramentaria (48.55%) was similar to those reported by (Crisan and Sands, 1978; Kalač, 2009) in the range 44.0-74.3% and 16.4-75%, respectively. Soluble sugars in edible mushrooms were highly regarded as biologically or medically active compounds and were used as functional food ingredients (Wasser and Weis, 1999). The content was 7.07% for this mushroom. Crude fat and ash contents here were 2.90 and 11.42%, respectively. These results were similar to those obtained by Crisan and Sands (1978), Yang et al. (2001) and Kalač (2009) in several edible mushrooms. From the results shown in Table-1, the macronutrient profile revealed that *Coprinopsis* atramentaria had rich sources of protein and fibre and had low amount of fat. This high protein and low fat characteristic of the edible wild mushrooms has been previously reported by many workers (Diez and Alvarez, 2001; Longvah and Deosthale, 1998).

Mineral elements

The metal content in the mushrooms are mainly affected by acidic and organic matter content of their ecosystem and soil (Gast et al., 1988). The uptake of metal ions in mushrooms is in many respects different from other plants. For this reason, the concentration variations of metals depend on mushroom species and their ecosystems (Seeger 1982). In general, most of the mushrooms studied had good amount of minerals including trace minerals. Potassium, Phosphorus, Sodium and Magnesium constitute about 56-70% of the total ash content of the mushrooms (Li and Chang, 1982), whereas potassium alone forms 45% of the total ash. Abou-Heilah et al. (1987) found that content of potassium and sodium in Coprinopsis atramentariawas 300 and 28.2 ppm, respectively. The concentration of sodium is relatively low and is of very great nutritional benefit to the consumer, a finding that has been corroborated by Vetter (2003). It is known that adequate iron in a diet is very important for decreasing the incidence of anaemia. The iron values of Coprinopsis atramentariaare in agreement with those reported in the literature (Sesli and Tuzen, 1999; Kalac, 2010). Copper concentrations in the accumulating mushroom species are usually100-300 mg/kg of dry matter, which is not considered as a health risk (Kalac, 2010; Kalac and Svoboda, 2000). Copper contents in mushrooms, higher than those in vegetables, should be considered as a nutritional source of the element. Nevertheless, for people, bioavailability from mushrooms was reported to be low, due to limited

Table 2: Antibacterial activity of different organic solvent extract of Coprinopsis atramentaria

Table 2. Antibacterial activity of different organic solvent extract of <i>Coprinopsis ulrumenturia</i>						
Test organisms	Hexane	Chloroform	Methanol	Ethyl	Acetate	
E. coli	2±0.01	4±0.12	8±0.01	6±0.03	7±0.16	
P. aeruginosa	7±0.13	4 ± 0.04	2±0.25	-	-	
S. typhi	2±0.14	13±0.19	8±0.13	-	±0.13	
S. aureus	3±0.23	5 ±0.16	16±0.12	12±0.32	11±0.17	
B. subtilis	12±0.18	7±013	8±0.29	-	-	
M. luteus	4±0.12	7±0.09	18±1.036	±0.02 14	±1.2	
$(7 - \dots - f(n))$						

(Zone of inhibition in -- mm); Activity expressed in millimetre (mm) concentration, (-) no activity.

absorption from the small intestine. Copper contents found in this study are in agreement with results reported in the literature (Sesli and Tuzen, 1999). Mushrooms were known as zinc accumulator and sporophore, substrate ratio for Zn ranges from 1 to 10 mg kg1 (Isiloglu *et al.*, 2001). The zincvalues of *Coprinopsis atramentaria* are in agreement with the reported values of the literature. Minerals generally in the diet are required for metabolic reactions, transmission of nerve impulses, rigid bone formation and regulation of water and salt balance among others.

Antibacterial activity

The increasing trend of resistance to the antibiotics in current use has drawn the attention of researchers to natural alternative treatments of bacterial infections as potential sources of novel antimicrobial agents. Gramnegative bacteria are reported to be resistant against most antibacterial agents as a result of the more complicated nature of their cell wall compared to gram-positive bacteria (Akinyemi et al., 2005; Ndukwe et al., 2007; Goyal et al., 2008). The antimicrobial activity of the extract Coprinopsis atramentaria showed prominent result against gram-positive organisms when compared with gram-negative organisms. The antimicrobial properties of the Coprinopsis atramentaria confirm previous studies that mushrooms possess antimicrobial effects (Hur et al., 2004; Sheena et al., 2003; Chang and Miles, 1989). Similar antimicrobial potentials have been observed in the culture extracts Agrocybe sp. (Arora, 1986; Mavoungou et al., 1987) and juice of L. Edodes (Kuznetosov et al., 2005).

Conclusion

On the whole, the mushrooms studied were found to be a good source of protein, fibre and minerals. The results from the present analysis allow a direct comparison of the chemical composition of Coprinopsis atramentaria with those of other mushrooms species. In conclusion, the edible mushroom Coprinopsis atramentaria consumed has essential medicinal properties. Boiling or cooking did not dilute or reduce the medicinal properties. Hence, it is necessary to identify the biological and pharmacological potential of mushrooms especially edible mushrooms that are collected indigenously and cultivated locally or sold in local and international market. The production and marketing of mushrooms and their products is vital for an economic importance. Therefore, it is also necessary to intensify research in identifying and isolating different varieties of mushrooms having nutraceutical and medicinal properties and to commercialize their production and marketing, which will boost the food industry and create employment to rural dwellers.

REFERENCES

- Abou-Heilah AN, MY Kasionalsim and AS Khaliel, 1987. Chemical composition of the fruiting bodies of *Agaricusbisporus*. Int J Exp Bot 47: 64-68.
- Akinyemi KO, O Oladapo, CE Okwara, CC Ibe and KA Fasure, 2005. Screening of crude extracts of six medicinal plants used in South-West Nigerian unorthodox medicine for anti-methicillin resistant *Staphylococcus aureus* activity. BMC Complement Altern Med 5:6.
- Aletor VA, 1990. Anti-Nutritional Factors in Some Nigerian Feedstuffs, Herbage by Products, Crops Resources and Browse Plants. A Monograph Prepared For the Presidential Task Force on Alternative Formulation of Live Stock Feed Products Development, Quality, Evaluation and Health Implications. Cabinet Office, Lagos, Nigeria.
- AOAC, 2000. Official Methods of Analysis. 15thed. Association of Official Analytical Chemists, Washington, DC, USA.
- Arora D, 1986. Mushroom demystified. Ten Spreed Press.
- Bender S, GT Loneigan, J Backhaus, CN Dumitrache-Anghel and WL Baker, 2001. The antibacterial activity of the edible and medicinal mushroom *Lentinusedodes* (Berk.) Sing Int J Med Mushrooms 3: 118.
- Boa F, 2004. Wild edible fungi: A global overview of their use and importance to people. FAO Press, pp: 37-78.
- Chang ST and PG Miles, 1989. Mushrooms: Cultivation, Nutritional Value, Medicinal Effects and Environmental Impacts, CRC Press, 4-6pp.
- Crisan EV and A Sands, 1978. Edible mushrooms: nutritional value. In. "The Biology and Cultivation of Edible Mushrooms" Ed. by ST Chang and WA Hayes, Academic Press, New York. pp: 137-165.
- Diez VA and A Alvarez, 2001. Compositional and nutritional studies on two wild edible mushrooms from northwest Spain, Food Chem, 75: 417-422.
- Fasidi IO, 1996. Studies on *Volvariella esculenta* mass singer, cultivation on agricultural wastes and proximate composition of stored mushrooms. Food Chem 55: 161-163.
- Gast CH, E Jansen, J Bierling and L Haanstra, 1988. Heavy metals in mushrooms and their relationship with soil characteristics. Chemosphere 17: 789-799.
- Goyal P, A Khanna, G Chauhan and P Kaushik, 2008. In vitro evaluation of crude extracts of *Catharanthusroseus* for potential antibacterial activity, Int J Green Pharm, 2: 176-181.
- Hur JM, Yang CH, Han SH, Lee SH, You YO and Park JC 2004. Antibacterial effect of *Phellinuslinteus* against methicillinresistant *Staphylococcusaureus*. Fitoterapia 73: 603-605.

- Isiloglu M, F Yilmaz and M Merdivan, 2001. Concentrations of trace elements in wild edible mushrooms. Food Chem, 73: 169-175.
- Jordan P, 2006. Field guide to edible mushrooms of Britain and Europe.New Holland publishers.
- Kalač P, 2009. Chemical composition and nutritional value of European species of wild growing mushrooms: a review. Food Chem, 113: 9-16.
- Kalač P, 2010. Trace element contents in European species of wild growing edible mushrooms: A review for the period 2000-2009. Food Chem 2:45. doi:10.1016/j.2010.
- Kalač P and L Svoboda, 2000. A review of trace element concentrations in edible mushrooms, Food Chem 69: 273-281.
- Karaman I, F Sahin, M Güllüce, H Ögütçü, M Sengül and A Adıgüzel, 2003. Antimicrobial activity of aqueous and methanol extracts of *Juniperusoxycedrus* LJ Ethnopharm, 85: 213-235.
- Kuznetsov OIU, Mil'kova EV, Sosnina AE and Sotnikova NIU, 2005. Antimicrobial action of *Lentinusedodes* juice on human microflora. *ZhMikrobiol Epidemiol Immunobiol* 1: 80-82.
- Li GSF and ST Chang, 1982. Nutritive value of *Volvariellavolvacea*. In. "Tropical Mushrooms-Biological Nature and Cultivation Methods".ed. by S T Chang and TH Quimio. Chinese University Press, Hong Kong. pp: 199-219.
- Longvah T and YG Deosthale, 1998. Compositional and nutritional studies on edible wild mushroom from northeast India. Food Chem, 63: 331-334.
- Mattila P, K Suonpaa and V Pilronen, 2000. Functional properties of edible mushrooms. Nutr, 16: 694-696.
- Mau JL, HC Lin and SF Song, 2002. Antioxidant properties of several specialty mushrooms. Food Res Int, 35: 519-526.
- Mavoungou H, M Porte and L Oddoux, 1987. Active anti tumoraled des myceliumsd'Agrocybedura, Mycoaciaudaet Phanerochaetelaevis. Ann Pharmaceutic Francaises, 45: 71-77.
- Moshi MJ, ZH Mbwambo, MC Kapingu, VH Mhozya and C Marwa, 2006. Antimicrobial and brine shrimp lethality of extracts of Terminaliamollislaws. Afr J Trad CAM 3: 1-10.
- Muhammad A, H Javid, TS Mohammad, KS Zabta, U Farman, B Ali, K Naeem, LK Abdul and W Takashi, 2010. Proximate and nutrient composition of medicinal plants of humid and sub-humid regions in North-west Pakistan. J Med Plant Res, 4: 339-345.
- Ndukwe IG, JO Amupitan, Y Isah and KS Adegoke, 2007. Phytochemicaland antimicrobial screening of the crude extracts from the root, stembark and leaves of *Vitellariaparadoxa*. Afr J Biotechnol 6: 1905-1909.
- Ndyetabura T, SL Lyantagaye and AM Mshandete, 2010. Antimicrobial activity of ethyl acetate extracts from edible Tanzanian *Coprinuscinereus* (schaeff) S

Grays. lat cultivated on grasses supplemented with cow dung manure. ARPN J AgricBiolSci 5(5).

- Oei P, 2010. Mushroom, Kluwers Publishers, Netherlands. 348pp.
- Okwulehie IC and EI Odunze, 2004. Evaluation of the nutritional value of some tropical edible mushrooms. J Sustain Agric Environ 6: 157-162.
- Okwulehie IC and ET Odunze, 2004. Evaluation of the myco-chemical and mineral composition of sometropical edible mushroom. J Sustain Agric Environ, 6: 63-70.
- Ouzouni PK, D Petridis, WD Koller and KA Riganakos, 2009. Nutritional value and metal content of wild edible mushrooms collected from West Macedonia and Epirus, Greece. Food Chem, 115: 1575-1580.
- Pearson D, 1976.Chemical Analysis of Foods.7th ed. J and AChurchill, London. pp: 34-36.
- Rai RD, 1994. Nutritional and medicinal values of mushrooms. In "Advances in Horticulture". ed by K L Chadha and SR Sharma. MalhotraPublishing House, New Delhi, pp: 537-551.
- Rai RD, 1997. Medicinal mushrooms. In."Advances in Mushroom Biology and Production".ed. by RD Rai, BL Dhar and RN Verma. MushroomSociety of India, NRCM, Solan, pp: 355-368.
- Rojas JJ, VJ Ochoa, SA Ocampo and JF Munoz, 2006. Screening for antimicrobial activity of ten Medicinal plants used in Colombian folkloric medicine: a possible alternative in the treatment of nonnosocomial infections. BMC Complement Altern Med 6: 2.
- Seeger R, 1982. Toxischeschwermetalle in Pilzen. Deutsche Apotheke Zeitschrift, 122: 1835-1844.
- Sheena N, A Ajith, TA Mathew and KK Janardhanan, 2003. Antibacterial activity of three Macro fungi Ganodermalucidum, Navesporusfloccose and Phellinusrimosus occurring in South India. Pharm Biol, 41: 564-567.
- Sesli E and M Tuzen, 1999. Levels of trace elements in the fruiting bodies of macro fungi growing in the East Black Sea region of Turkey. Food Chem, 65: 453-460.
- Van SPJ, 1967. Development of a comprehensive system of food analysis and its application to forages. J Am Sci, 26: 119-128.
- Vetter J, 2003. Data on sodium content of common edible mushrooms. Food Chem, 81: 589-593.
- Whistler R and M Wolfrom, 1962. Methods in Carbohydrate Chemistry, Vol, 1. Academic Press, New York.
- Wasser SP and AL Weis, 1999. Medicinal properties of substances occurring in higher Basidiomycetes mushrooms: current perspective (review). Int J Med Mushrooms, 1: 31-62.
- Yang JH, HC Lin and JL Mau, 2001. Non-volatile taste components of several speciality mushrooms. Food Chem, 72: 465-471.